

## Microdeletion of *LIT1* in Familial Beckwith-Wiedemann Syndrome

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Beckwith-Wiedemann syndrome (BWS), which causes prenatal overgrowth, midline abdominal wall defects, macroglossia, and embryonal tumors, is a model for understanding the relationship between genomic imprinting, human development, and cancer. The causes are heterogeneous, involving multiple genes on 11p15 and including infrequent mutation of *p57<sup>KIP2</sup>* or loss of imprinting of either of two imprinted gene domains on 11p15: *LIT1*, which is near *p57<sup>KIP2</sup>*, or *H19/IGF2*. Unlike Prader-Willi and Angelman syndromes, no chromosomal deletions have yet been identified. Here we report a microdeletion including the entire *LIT1* gene, providing genetic confirmation of the importance of this gene region in BWS. When inherited maternally, the deletion causes BWS with silencing of *p57<sup>KIP2</sup>*, indicating deletion of an element important for the regulation of *p57<sup>KIP2</sup>* expression. When inherited paternally, there is no phenotype, suggesting that the *LIT1* RNA itself is not necessary for normal development in humans.

### Introduction

Beckwith-Wiedemann syndrome (BWS [MIM 130650]) rarely involves mutation of the imprinted cyclin-dependent kinase inhibitor gene *p57<sup>KIP2</sup>* (MIM 600856), which is normally expressed from the maternal allele (Matsuo et al. 1996; Lee et al. 1997; O'Keefe et al. 1997). However, 40% of patients with BWS show loss of imprinting (LOI) of *LIT1* (MIM 604115), an antisense RNA within the *KvLQT1* gene (MIM 192500) and 220 kb telomeric to *p57<sup>KIP2</sup>* on chromosome 11p15 (Lee et al. 1999; Smilnich et al. 1999). An additional 15% of patients show LOI of *IGF2* (MIM 147470), located in a second imprinted subdomain 280 kb telomeric to *KvLQT1* (Weksberg et al. 1993; Steenman et al. 1994). LOI of *LIT1* involves aberrant hypomethylation and activation of the normally silent maternal allele. The target of LOI of *LIT1* presumably is *p57<sup>KIP2</sup>*, although it is not clear whether *LIT1* serves as a transcriptional repressor of *p57<sup>KIP2</sup>* or as an insulator separating *p57<sup>KIP2</sup>* from its enhancer. Deletion of the differentially methylated region (DMR) leads to activation of the paternal allele of *p57<sup>KIP2</sup>*, as well as those of *Tssc3*, *Tssc5*, *Kcnq1*, *Tssc4*, and *Ascl2* (Fitzpatrick et al. 2002). Similarly, targeted deletion of the *LIT1* CpG island on a human paternal

chromosome in chicken DT40 cells leads to activation of the paternal allele of *KvLqt1* and *p57<sup>KIP2</sup>* (Horike et al. 2000). Presumably, the DMR contains elements important for silencing on the paternal allele. An engineered translocation in the mouse, disrupting the imprinted cluster with a targeted translocation between *p57<sup>KIP2</sup>* and *KvLqt1*, results in loss of expression of the maternal allele of *p57<sup>KIP2</sup>*, *Tssc3*, and *Tssc5* (Cleary et al. 2001), suggesting that there are also elements within or proximal to *LIT1* important for gene activation on the maternal allele. It should be noted, however, that there is no mouse model involving *LIT1* leading to BWS.

We sought to identify patients with BWS who have microdeletions on 11p15, since similar studies had clarified the role of imprinting regulatory elements in the Prader Willi/Angelman syndrome gene region of 15q11-15q13 (Buiting et al. 1995). To screen for *LIT1* deletions, we used real-time quantitative PCR of genomic DNA for copy number analysis, normalizing to a reference locus, and combined these efforts with FISH of metaphase and interphase cells.

### Subjects and Methods

#### Subjects

Patients with BWS were identified as part of a BWS registry first established in 1994. Participating families provided detailed clinical information and written informed consent approved by the respective institutional review boards. BWS in the registry is defined by a clinical diagnosis by a experienced physician but requires at least two of the five most common features: (1) macroglossia,

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(2) birth weight and length >90th percentile, (3) hypoglycemia in the 1st mo of life, (4) ear creases or ear pits, and (5) midline abdominal wall defects.

#### Real-Time Karyotyping

Genomic DNA was prepared from peripheral-blood lymphocytes by standard proteinase K digestion and phenol extraction (Cui et al. 1998). Genomic copy numbers of sequences within *K<sub>v</sub>LQT1* and flanking genes were measured using quantitative real-time PCR amplification with Taqman probes. Reactions were performed using 1 × Taqman master mix (ABI), 300 nM each primer, 200 nM probe, and 10 ng of peripheral white blood cell genomic DNA in a total volume of 25 μl. All reactions were performed in triplicate. For each sequence amplified, triplicate reactions of a no template control and a standard curve were amplified at the same time. The standard curve was prepared from normal control genomic DNA at 100 × (100 ng per reaction), 10 × (10 ng per reaction), and 1 × (1 ng per reaction). Reactions were run on an ABI 7700 instrument under standard conditions (50°C for 2 min; 95°C for 10 min; 95°C for 15 s, 60°C for 1 min for 40 cycles). All triplicate cycle threshold (Ct) values were checked to ensure that they were within 1 Ct of each other. The log input amount of the standard curve was plotted versus the output Ct values; only amplifications that produced a slope of -3.3 to -3.8 were accepted as quantitative. The log input amount of each sample was calculated according to the formula  $(Ct - b)/m$ , where  $b$  is the Y-intercept, and  $m$  is the slope. The log input amount was converted to input amount according to the formula  $10^{(\log \text{ input amount})}$ , and triplicate input amounts were averaged for each sample. The average input amount of each sequence amplified was normalized to the average input amount of peripheral myelin protein 22 (*PMP22*); primer and probe sequences for *PMP22* were reported by Wilke et al. (2000). *PMP22* is located on chromosome 17 and is involved in Charcot-Marie-Tooth syndrome, a sensory and motor neuropathy disease unrelated to BWS. All primers are listed in the appendix (online only).

#### FISH

To confirm the deletion localization in lymphoblast cells derived from patients II-2 and III-4, two-color FISH was performed using several BAC and PAC clones, including *K<sub>v</sub>LQT1* genomic DNA as a probe. The probes were labeled with digoxigenin-11-UTP (Roche) and biotin-16-dUTP (Roche), respectively, by nick translation. Probes were purified by ethanol precipitation, dissolved in 5 μl of formamide, mixed, and denatured. Hybridization solution (BSA [Roche]:10 × SSC:50% dextran sulfate [Sigma], 1:2:2) and labeled probes were mixed 1:1,

dropped onto denatured chromosomes, covered with parafilm, and incubated at 37°C for 15 h in a humidified chamber. After hybridization, the slides were washed sequentially at 37°C in 50% formamide/2 × SSC, 2 × SSC, 1 × SSC for 15 min each and once in 4 × SSC for 5 min. The slides were immersed in 70 μl of 3 μg/ml fluorescein isothiocyanate (FITC)-avidin (Vector Laboratories), and 1.6 μg/ml anti-digoxigenin-rhodamine (Roche), 4 × SSC, and 1% BSA for 45 min at 37°C. The slides were then washed for 5 min each in 4 × SSC, 4 × SSC containing 0.05% Triton X-100, and then 4 × SSC. The slides were incubated with 70 μl of 5 μg/ml biotinylated anti-avidin (Vector Laboratories), 4 × SSC, and 1% BSA at 37°C for 60 min. After washing, another layer of FITC-avidin was added for amplification. The slides were washed and mounted in antifade solution (1% diazabicyclooctane [Sigma] in glycerol with 10% PBS) containing 0.2 μg/ml 4',6'-diamidino-2-phenylindole (Sigma) and 100 μg/ml p-phenylenediamine (Sigma). Signals were observed with a Nikon fluorescence microscope. One hundred nuclei and metaphases were analyzed.

#### Real-Time Quantitative Analysis of Gene Expression

RNA isolated from primary lymphocytes was converted to cDNA through use of the Superscript II kit (Invitrogen) according to the manufacturer's protocol. A standard curve was constructed from pooled normal control cDNA. Reactions were performed using 1 × Taqman master mix (Applied Biosystems), 270 nM each primer, and 60 nM probe in a total volume of 25 μl. *GUS* reactions were performed using 24 ng of original input RNA, and *p57<sup>KIP2</sup>* reactions were performed using 192 ng of original input RNA. All reactions were performed in triplicate, including, for each sequence amplified, a no-template control. The RT-negative reactions were used as template for one *p57<sup>KIP2</sup>* reaction, to ensure that no amplification occurred. Reactions were run on an ABI 7700 instrument according to standard conditions (50°C for 2 min; 95°C for 10 min; 95°C for 15 s, 60°C for 1 min for 40 cycles). All triplicate Ct values were checked to ensure that they were within 1 Ct of each other. For *p57<sup>KIP2</sup>*, the average slope of the standard curve was 3.73, and for *GUS*, the average slope of the standard curve was 3.68; both of these are within the desired quantitative range. Data analysis converting triplicate Ct values into average input amount ratios was performed as described above in the "Real-Time Karyotyping" subsection. The SD was, on average, 23% of the ratio value. A human endogenous control plate (Applied Biosystems) was used, according to the manufacturer's instructions, to choose a housekeeping gene for expression normalization in primary lymphocytes.

## Results

We used real-time quantitative PCR of genomic DNA to screen for microdeletions in BWS. We examined 34 patient samples by this approach. Of these, two patients had familial BWS, each with at least one fully affected sib. We detected a microdeletion in one of the familial cases, as described in detail below. The other familial case showed evidence of a complex rearrangement (not discussed here).

At the time of ascertainment, patient III-4 (fig. 1A) was a 6-year-old boy with hemihypertrophy with asymmetric enlargement of the right leg and arm, hypospadias, cleft palate, midline abdominal wall defect (diastasis recti and umbilical hernia), characteristic ear pits and creases, undescended testes, and macroglossia requiring surgery. Two siblings also had BWS: a sister (patient III-1, fig. 1A) with prenatal overgrowth, ear creases and pits, and macroglossia requiring surgery; and a brother (patient III-2, fig. 1A), stillborn at 28 wk, with prenatal overgrowth and macroglossia. The father was 37 years old and the mother 36 years old at the proband's birth, and both parents showed no signs of BWS in childhood or later.

The deletion was mapped by real-time karyotyping with probes spanning *K<sub>v</sub>LQT1* (fig. 2A). The deleted region of ~250 kb removes all of *LIT1* and is within *K<sub>v</sub>LQT1*; the centromeric breakpoint is located between exons 1b and 1c, and the telomeric breakpoint is located between exons 11 and 14 (*K<sub>v</sub>LQT1* exon numbering from GenBank sequence AJ006345). This deletion was detected in individuals II-2 and III-4; II-2 is the mother of III-4 (fig. 1A), indicating that it had been transmitted from the mother to the proband and likely to the other two deceased sibs with BWS as well.

The presence and localization of the deletion was independently confirmed cytogenetically through use of two-color interphase and metaphase FISH with BAC and PAC probes spanning *K<sub>v</sub>LQT1* (fig. 2B). PAC probes AC002403 and U90095, spanning *K<sub>v</sub>LQT1* from approximately exon 2a to exon 11, generated one chromosomal signal in a majority of interphase and metaphase nuclei in diploid lymphoblast cells derived from patients II-2 and III-4, whereas flanking probes AC003693 (PAC) and AC013791 (BAC) generated two chromosomal signals (fig. 2B).

To determine the identity of the parental allele carrying the deletion, we performed haplotype analysis, using microsatellite markers across 11p15 (fig. A [online only]). Through use of these data, haplotype analysis of the available individuals placed the deletion on the "ACIL" haplotype (fig. 1A). Individual II-2 inherited the chromosome carrying this haplotype from her father, individual I-2, and passed this haplotype to her son, individual III-4, indicating that the deletion was present on

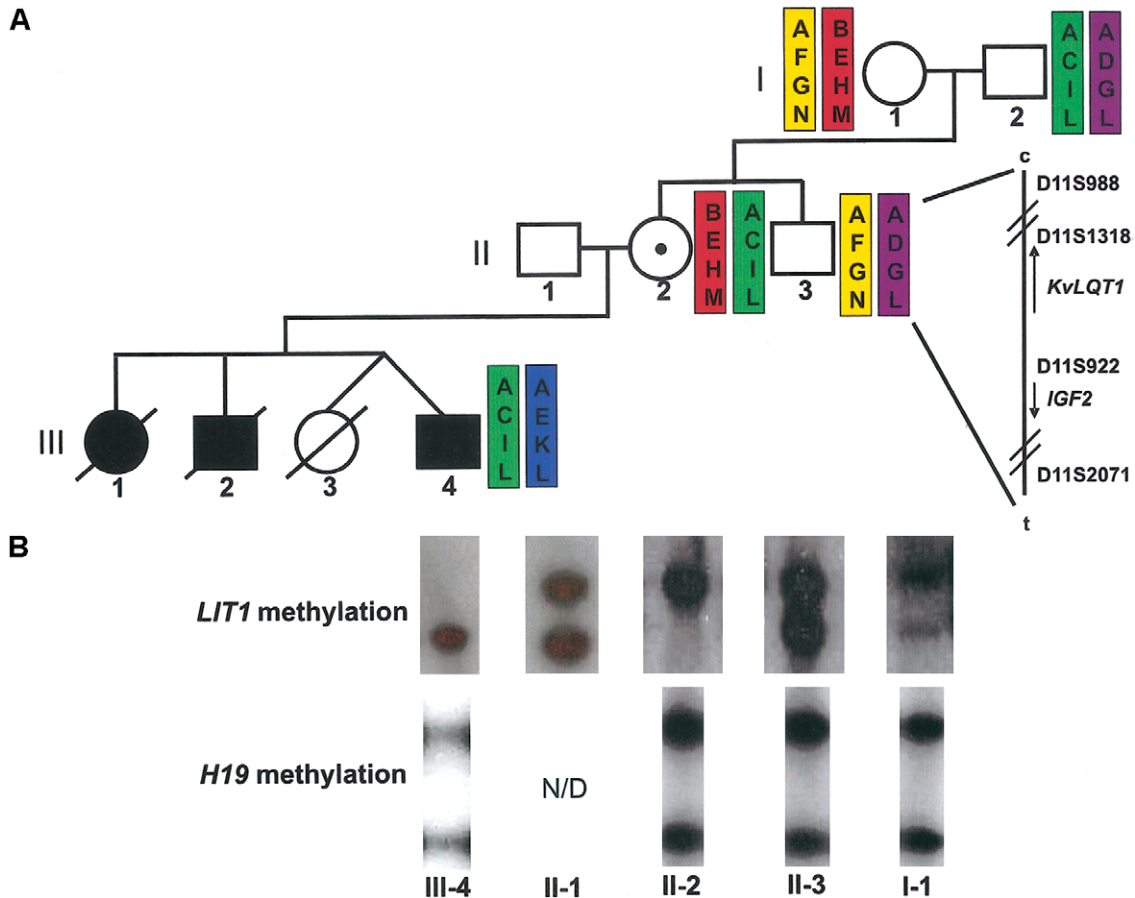
a paternal chromosome in individual II-2 and on a maternal chromosome in individual III-4. Methylation analysis of the *LIT1* DMR by methyl-sensitive Southern blotting (fig. 1B) was used to confirm the parental origin and indicated the absence of an unmethylated paternal band in individual II-2 and the absence of a methylated maternal band in individual III-4. Methylation analysis of the *H19* DMR by methyl-sensitive Southern blotting (fig. 1B) was used to confirm that the deletion had no effect on the distal, independently regulated *H19/IGF2* cluster and indicated the normal presence of both parental bands in all tested individuals.

The patient's mother inherited the deletion from her father, and thus we could compare the effect on gene expression of a maternally inherited deletion with that of a paternally inherited deletion. If the deletion removed a *p57<sup>KIP2</sup>* enhancer, then the patient would be expected to show reduced expression with no change in expression in the mother. In contrast, if *LIT1* acts as a silencer—that is, as a repressor of *p57<sup>KIP2</sup>* over a distance—then the paternally inherited allele would be expected to be activated. To determine the relationship between the microdeletion and expression of *p57<sup>KIP2</sup>*, we performed real-time quantitative RT-PCR of *p57<sup>KIP2</sup>* (fig. 2C). We could not measure allele-specific expression of *p57<sup>KIP2</sup>* directly, since there were no polymorphisms in the exons, introns, or promoter of *p57<sup>KIP2</sup>*. *TSSC3* and *TSSC5* are not consistently imprinted in humans postnatally, so the effect on imprinting of more-distant genes could not be assessed.

The expression level of *p57<sup>KIP2</sup>* was measured in peripheral blood lymphocyte RNA from individuals III-4 and II-2 and compared with normal age-matched controls. These data indicated that *p57<sup>KIP2</sup>* expression is reduced in the proband (relative expression level 1.2, versus a mean  $\pm$  SD of  $7.9 \pm 3.8$  in age-matched controls [fig. 2C]). In contrast, expression of *p57<sup>KIP2</sup>* was normal in the patient's mother, compared with that in age-matched controls (fig. 2C). Therefore, the microdeletion decreased *p57<sup>KIP2</sup>* expression when inherited maternally but had no effect when inherited paternally, suggesting a mechanism of enhancer deletion. In addition, *LIT1* itself showed normal expression in the proband but no detectable expression in the proband's mother (fig. 2C), consistent with loss of her paternally inherited copy, which is normally the only active allele. Since the mother had no growth or developmental abnormalities, the absence of *LIT1* RNA had no discernible phenotypic effect on her.

## Discussion

In summary, we have described a microdeletion on 11p15 in two related individuals, one in whom the deletion is maternally inherited and the other in whom the

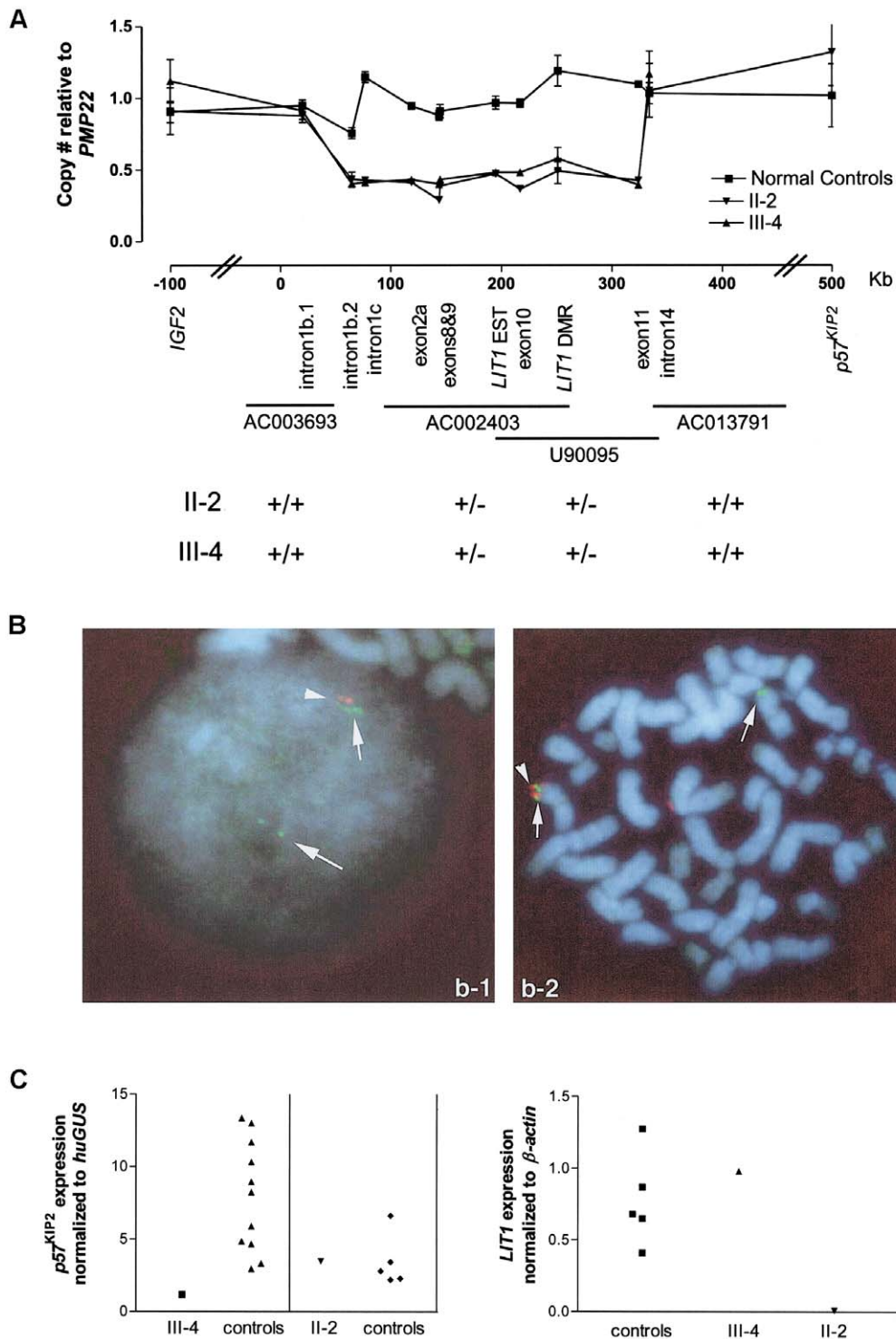


**Figure 1** Haplotype and methylation analysis of *LIT1* microdeletion. *A*, Haplotype mapping using 11p15 microsatellite markers D11S988, D11S1318, D11S922, D11S2071, which were amplified using FAM-labeled primers and detected using an ABI 3100 capillary electrophoresis instrument. Haplotypes were constructed using peak size data (see fig. A [online only]). The haplotypes of individuals I-1 and II-3 are inferred. *B*, Methylation at *LIT1* and *H19*, assayed by methyl-sensitive Southern blotting using genomic DNA from the indicated individuals. For *LIT1*, the upper band (6.0 kb) is methylated and represents the maternal allele, and the lower band (4.2 kb) is unmethylated and represents the paternal allele. For *H19*, the upper band (1.8 kb) is methylated and represents the paternal allele, and the lower band (1.0 kb) is unmethylated and represents the maternal allele. *LIT1* shows gain of methylation in II-2 and loss of methylation in III-4. *H19* is unaffected.

deletion is paternally inherited. Although the mechanism of regulation of the imprinted cluster of the genes *IGF2* and *H19* has been well studied and related to the BWS phenotype, the regulation and disease-causing mechanism of the imprinted cluster of the genes *p57<sup>KIP2</sup>*, *KvLQT1*, *LIT1*, *TSSC3*, and *TSSC5* is less well understood. In humans, it is clear that inactivating mutations in *p57<sup>KIP2</sup>* cause BWS, and LOI and aberrant methylation of *LIT1* has been associated with a third of all BWS cases. A number of studies in mice have indirectly shown that *LIT1* plays a role in regulating imprinted expression of the nearby genes *p57<sup>KIP2</sup>*, *TSSC3*, and *TSSC5*. In mice, disruption of the imprinted cluster with a targeted translocation between *p57<sup>KIP2</sup>* and *KvLQT1* results in loss of expression and loss of imprinting of the genes *p57<sup>KIP2</sup>*, *TSSC3*, and *TSSC5* (Cleary et al. 2001), indicating that *KvLQT1* harbors important regulatory elements. How-

ever, knockout of the DMR of *LIT1* in mice results in derepression of nearby imprinted genes and a micro-somic phenotype when inherited paternally, although there is no affect on growth or imprinted gene expression when inherited maternally (Fitzpatrick et al. 2002).

Here, we provide the first human genetic evidence that the *LIT1* domain contains regulatory elements that play a mechanistic role in the BWS phenotype and regulates the imprinting and expression of nearby imprinted genes. This microdeletion provides genetic confirmation of the importance of regulation of *p57<sup>KIP2</sup>* at a distance in the pathogenesis of BWS. The model that best fits these data is that, in the case of the maternal deletion, *p57<sup>KIP2</sup>* expression is lost because of loss of an enhancer within or telomeric to *LIT1*, leading to BWS, which supports the prediction of an engineered translocation in the mouse (Cleary et al. 2001). Consistent



**Figure 2** Deletion of *LIT1* and altered expression of *p57<sup>KIP2</sup>*. *A*, Genomic copy number of sequences in *KvLQT1* in individuals II-2 and III-4 and in four normal control individuals. Genomic DNA was amplified in the presence of a Taqman probe and normalized to *PMP22* on chromosome 17. The X-axis indicates *KvLQT1* genomic sequence (GenBank accession number AJ006345), and the Y-axis indicates the relative copy number of each probed sequence. See the appendix (online only) for primer and probe sequences. *B*, Two-color FISH using PAC probes in lymphoblastoid nuclei from patient III-4. Probe AC003693 (blue) generated two chromosome signals in interphase (left) and metaphase (right) nuclei, whereas probe U90095 (red) generated one chromosome signal in interphase and metaphase nuclei. A pair of spots was scored as one signal, since it represents the two sister chromatids after S phase. Interphase nuclei and chromosomes were counterstained with DAPI. One hundred interphase and metaphase nuclei were analyzed, 90 and 85 of which, respectively, showed similar patterns to those described above. *LIT1* is deleted in individuals II-2 and III-4. *C*, Real-time mRNA expression analysis of *p57<sup>KIP2</sup>* and *LIT1* in deletion carriers. Values represented are the average input amount of *p57<sup>KIP2</sup>*, normalized to the average input amount of the  $\beta$ -glucuronidase (*GUS*) gene in triplicate samples, compared with age-matched normal lymphocyte controls; and the average input amount of *LIT1*, normalized to the average input amount of the  $\beta$ -actin gene in triplicate samples, compared with normal lymphoblastoid controls. The housekeeping gene chosen for normalization in cases was the gene with the least variance among eight control samples. *p57<sup>KIP2</sup>* expression is decreased in the proband (III-4), and *LIT1* expression is absent in the mother (II-2).

with this idea, Diaz-Meyer et al. (2003) found incomplete silencing of  $p57^{KIP2}$  in several patients with BWS who had LOI of *LIT1*, compared with patients without BWS, although patients with BWS who didn't have LOI were not examined in that study.

In addition, absence of an *LIT1* transcript, as found in the proband's mother, had no phenotypic consequence, indicating that the *LIT1* RNA has no significant physiological function when inherited paternally. The deletion in this family is also of practical importance that should be brought to the attention of clinical geneticists performing molecular diagnosis of BWS. Several centers worldwide are performing methylation analysis for BWS, the results of which will be abnormal for these individuals. However, unlike LOI of *LIT1*, in which the recurrence risk and transmissibility are unknown, definitive counseling can be provided to patients with the deletion by using methylation analysis and either digital karyotyping or FISH with the appropriate probes. A mother carrying the deletion would have a 50% risk of transmission to subsequent offspring, and similar counseling could be provided to the probands as well.

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## Electronic-Database Information

The accession number and URLs for data presented herein are as follows:

GenBank, <http://www.ncbi.nlm.nih.gov/Genbank/> (for *KvLQT1* genomic sequence [accession number AJ006345])  
 Online Mendelian Inheritance in Man (OMIM), <http://www.ncbi.nlm.nih.gov/Omim/> (for BWS,  $p57^{KIP2}$ , *LIT1*, *KvLQT1*, and *IGF2*)

## References

Buiting K, Saitoh S, Gross S, Dittrich B, Schwartz S, Nicholls RD, Horsthemke B (1995) Inherited microdeletions in the Angelman and Prader-Willi syndromes define an imprinting centre on human chromosome 15. *Nat Genet* 9:395–400  
 Cleary MA, van Raamsdonk CD, Levorse J, Zheng B, Bradley A, Tilghman SM (2001) Disruption of an imprinted gene

cluster by a targeted chromosomal translocation in mice. *Nat Genet* 29:78–82  
 Cui H, Horon IL, Ohlsson R, Hamilton SR, Feinberg AP (1998) Loss of imprinting in normal tissue of colorectal cancer patients with microsatellite instability. *Nat Med* 4:1276–1280  
 Diaz-Meyer N, Day CD, Khatod K, Maher ER, Cooper W, Reik W, Junien C, Graham G, Algar E, Der Kaloustian VM, Higgins MD (2003) Silencing of *CDKN1C* ( $p57^{KIP2}$ ) is associated with hypomethylation at *KvDMR1* in Beckwith-Wiedemann syndrome. *J Med Genet* 40:797–801  
 Fitzpatrick GV, Soloway PD, Higgins MJ (2002) Regional loss of imprinting and growth deficiency in mice with a targeted deletion of *KvDMR1*. *Nat Genet* 32:426–431  
 Horike S, Mitsuya K, Meguro M, Kotobuki N, Kashiwagi A, Notsu T, Schulz TC, Shirayoshi Y, Oshimura M (2000) Targeted disruption of the human *LIT1* locus defines a putative imprinting control element playing an essential role in Beckwith-Wiedemann syndrome. *Hum Mol Genet* 9:2075–2083  
 Lee MP, DeBaun MR, Mitsuya K, Galonek HL, Brandenburg S, Oshimura M, Feinberg AP (1999) Loss of imprinting of a paternally expressed transcript, with antisense orientation to *KvLQT1*, occurs frequently in Beckwith-Wiedemann syndrome and is independent of insulin-like growth factor II imprinting. *Proc Natl Acad Sci USA* 96:5203–5208  
 Lee MP, DeBaun M, Randhawa G, Reichard BA, Elledge SJ, Feinberg AP (1997) Low frequency of  $p57^{KIP2}$  mutation in Beckwith-Wiedemann syndrome. *Am J Hum Genet* 61:304–309  
 Matsuoka S, Thompson JS, Edwards MC, Bartletta JM, Grundy P, Kalikin LM, Harper JW, Elledge SJ, Feinberg AP (1996) Imprinting of the gene encoding a human cyclin-dependent kinase inhibitor,  $p57^{KIP2}$ , on chromosome 11p15. *Proc Natl Acad Sci USA* 93:3026–3030  
 O'Keefe D, Dao D, Zhao L, Sanderson R, Warburton D, Weiss L, Anyane-Yeboah K, Tycko B (1997) Coding mutations in  $p57^{KIP2}$  are present in some cases of Beckwith-Wiedemann syndrome but are rare or absent in Wilms' tumors. *Am J Hum Genet* 61:295–303  
 Smilnich NJ, Day CD, Fitzpatrick GV, Caldwell GM, Lossie AC, Cooper PR, Smallwood AC, Joyce JA, Schofield PN, Reik W, Nicholls RD, Weksberg R, Driscoll DJ, Maher ER, Shows TB, Higgins MJ (1999) A maternally methylated CpG island in *KvLQT1* is associated with an antisense paternal transcript and loss of imprinting in Beckwith-Wiedemann syndrome. *Proc Natl Acad Sci USA* 96:8064–8069  
 Steenman MJ, Rainier S, Dobry CJ, Grundy P, Horon IL, Feinberg AP (1994) Loss of imprinting of *IGF2* is linked to reduced expression and abnormal methylation of *H19* in Wilms' tumor. *Nat Genet* 7:433–439  
 Weksberg R, Shen DR, Fei YL, Song QL, Squire J (1993) Disruption of insulin-like growth factor 2 imprinting in Beckwith-Wiedemann syndrome. *Nat Genet* 5:143–150  
 Wilke K, Duman B, Horst J (2000) Diagnosis of haploidy and triploidy based on measurement of gene copy number by real-time PCR. *Hum Mutat* 16:431–436